

# Understanding Nanoparticle Transport in Human Vasculature Through Large-scale Fluid-Structure Interaction Simulations

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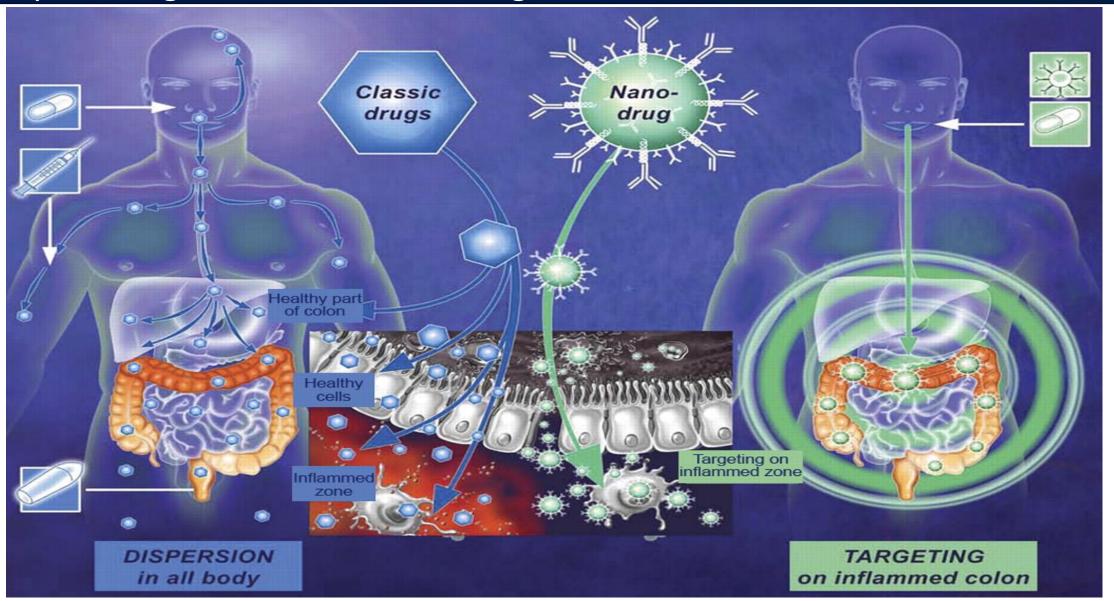
Collaborators: Wing Kam Liu, Northwestern University; Martin Kroger, ETH Zurich; Dean Ho, UCLA; Paolo Decuzzi, Italian Institute of Technology; Mu-Ping Nieh, Jessica Rouge, Mei Wei, Xiuling Lu, UConn; Lucy Zhang, RPI Students: Dr. Huilin Ye (Cadence), Dr. Zhiqiang Shen (ORNL), Alessandro Fisher, William Baker (REU), Jeffrey Ge (REU)

Palabos Online Seminar Series, September 1st, 5:00 pm CET, 2021

- ☐ Background
- ☐ Computational framework
- Validation and Verification
- ☐ Results
- Anomalous vascular dynamics of nanoworms within blood Flow
- Adhesion effect on margination of elastic microparticles
- Motion of particle under external magnetic field
- ☐ Conclusion and Ongoing work

# Nanomedicine: a promising method to deliver drugs to diseased





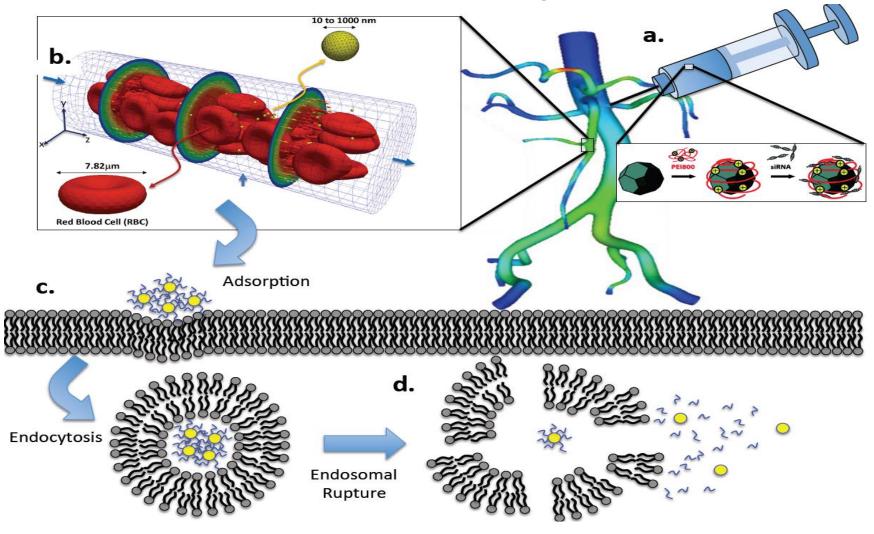
Am. J. Physiol. Gastrointest. Liver Physiol. 300.3 (2011): G371-G383.



# Computational design of efficient drug carriers



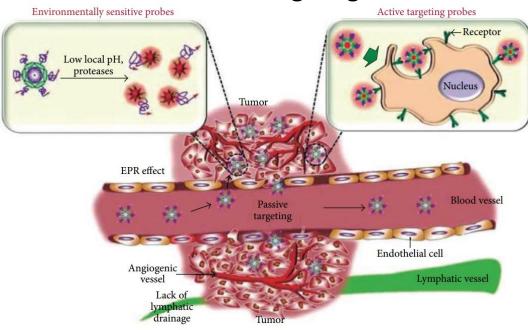
Goal: Design nanoparticle-based drug carriers for efficient and targeted drug delivery via predictive multiscale modeling



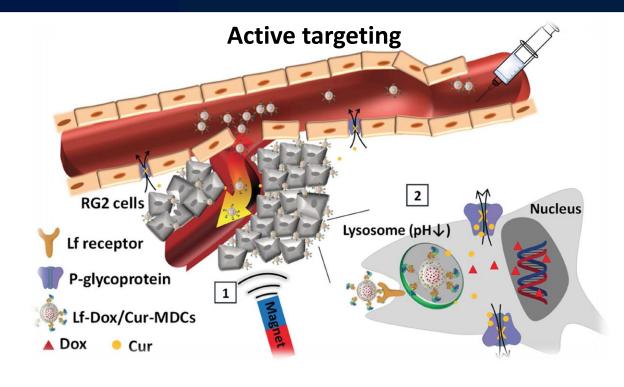
## **Background Information**

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#### **Passive targeting**



- **☐** Life journey
- Transport and circulation with RBCs, WBCs, and others in normal vascular network
- Firm adhesion to the vessel wall near tumor site
- Diffusion into tumor site through 'leaky' vessel walls
- > Recognition and uptake by tumor cells



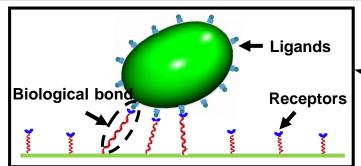
- **☐** Intrinsic properties
- Size
- > Shape
- > Stiffness
- > Surface functionality

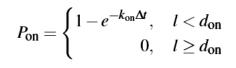
- External triggering
- Electrical
- Magnetical
- > Chemical
- > Optical

Ye, Huilin, et al. Proc. R. Soc. A (2018)

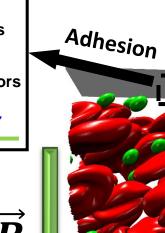
# **Computational Model**

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$$P_{ ext{off}} = egin{cases} 1 - e^{-k_{ ext{off}}\Delta t}, & l < d_{ ext{off}} \ 0, & l \geq d_{ ext{off}} \end{cases}$$



#### Incompressible Flow

$$\nabla \cdot \boldsymbol{v} = 0$$

$$\rho(\frac{\partial \boldsymbol{v}}{\partial t} + \boldsymbol{v} \cdot \nabla \boldsymbol{v}) = -\nabla p + \mu \nabla^2 \boldsymbol{v} + \boldsymbol{f}$$

#### Lattice Boltzmann Method

$$f_i(\mathbf{x} + \mathbf{e}_i \Delta t, t + \Delta t) = f_i(\mathbf{x}, t) - \frac{1}{\tau} (f_i(\mathbf{x}, t) - f_i^{eq}(\mathbf{x}, t))$$



$$u_i^{\rm s}(\mathbf{X}^{\rm s},t) = \int_{\Omega} u_i(\mathbf{x},t) \delta(\mathbf{x} - \mathbf{x}^{\rm s}(\mathbf{X}^{\rm s},t)) d\Omega$$

$$f_i^{FSI,s}(\mathbf{X},t) = \int_{\Omega^s} F_i^{FSI,s}(\mathbf{X}^s,t) \delta(\mathbf{X} - \mathbf{X}^s(\mathbf{X}^s,t)) d\Omega$$

Red blood cells and particles

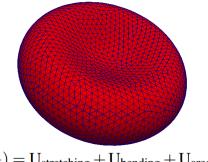
Plasma

# $\mathbf{m} = \frac{V_c \chi}{\mu^*} \mathbf{B}$

$$\mathbf{F}^B = (\mathbf{m} \cdot \nabla)\mathbf{B}$$

$$\mathbf{F}_{ij}^{B} = \frac{3\mu^{*}}{4\pi r^{4}} \left( 2(\mathbf{m} \cdot \mathbf{e}_{ij})\mathbf{m} - (5(\mathbf{m} \cdot \mathbf{e}_{ij})^{2} - m^{2})\mathbf{e}_{ij} \right)$$

#### Coarse-grained Method



$$U(\{\boldsymbol{x}_i\}) = U_{\text{stretching}} + U_{\text{bending}} + U_{\text{area}} + U_{\text{volume}} \quad U_{\text{volume}} = \frac{k_v(V - V_0)}{2V_0}$$

$$U_{WLC} = \frac{k_B T l_m}{4p} \frac{3x^2 - 2x^3}{1 - x}, \ U_{POW} = \frac{k_p}{l}$$

$$U_{bending} = \sum_{k \in 1...N_s} k_b [1 - \cos(\theta_k - \theta_0)]$$

$$U_{area} = \sum_{k=1..N_{t}} \frac{k_{d}(A_{k} - A_{k0})^{2}}{2A_{k0}} + \frac{k_{a}(A_{t} - A_{t0})^{2}}{2A_{t}}$$

$$U_{\text{volume}} = \frac{k_{\text{v}}(V - V_0)}{2V_0}$$

### **Computational Model-Fluid Flow**



The lattice-Boltzmann algorithm is a popular method used to simulate the hydrodynamics of complex fluids on a grid. As this method requires only nearest neighbor grid point information, it is very straightforward to implement, and is ideally suited for large-scale parallel applications.

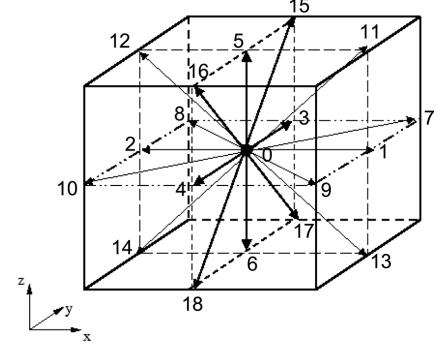
Navier-Stokes equations with thermal noise (Landau & Lifshitz):

$$(\partial_t + \partial_\alpha u_\alpha)\rho = 0$$

$$\rho(\partial_t + u_\alpha \partial_\alpha) u_\beta = -\partial_\alpha (P_{\beta\alpha} + s_{\beta\alpha}) + \partial_\alpha (\eta_{\alpha\beta\gamma\nu} \partial_\gamma u_\nu) + F_\beta$$

 Note that the thermal noise appears in the stress tensor so will conserve mass and momentum. It should also obey the fluctuationdissipation theorem (Landau & Lifshitz):

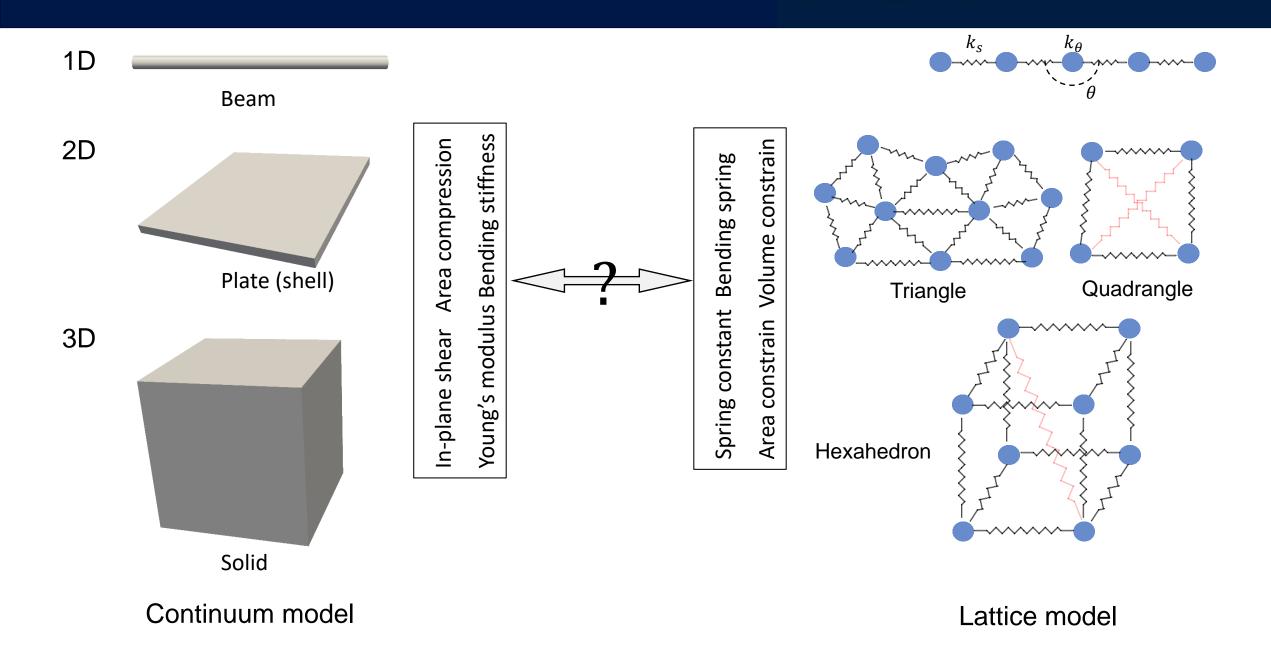
$$\langle s_{\alpha\beta}(\mathbf{r},t)s_{\gamma\nu}(\mathbf{r}',t')\rangle = 2k_BT\eta_{\alpha\beta\gamma\nu}\delta(\mathbf{r}-\mathbf{r}')\delta(t-t')$$



Chen, Shiyi, and Gary D. Doolen. "Lattice Boltzmann method for fluid flows." *Annual review of fluid mechanics* 30, no. 1 (1998): 329-364.

#### **Structure: Lattice Model**

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#### Structure: shell/membrane

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#### Shell (Red blood cell and sphere)

$$U(\{\mathbf{x}_i\}) = U_{stretching} + U_{bending} + U_{area} + U_{volume}$$

In-plane stretching (bond)

$$U_{WLC} = \frac{k_B T l_m}{4p} \frac{3x^2 - 2x^3}{1 - x}, \ U_{POW} = \frac{k_p}{l}$$

Out-plane bending (dihedral)

$$U_{bending} = \sum_{k \in 1...N_s} k_b [1 - \cos(\theta_k - \theta_0)]$$

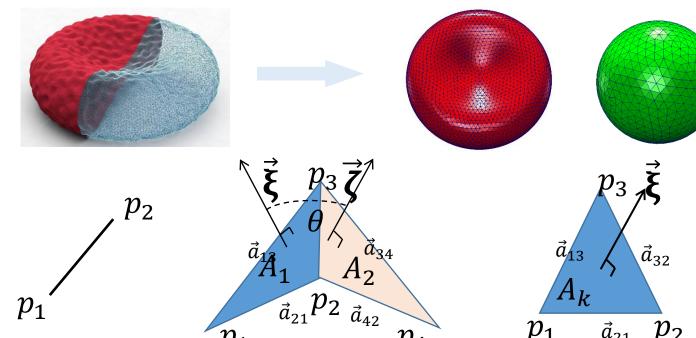
Area incompressibility of the lipid bilayer

$$U_{area} = \sum_{k=1...N_t} \frac{k_d (A_k - A_{k0})^2}{2A_{k0}} + \frac{k_a (A_t - A_{t0})^2}{2A_t}$$

Incompressibility of the inner cytosol

$$U_{\text{volume}} = \frac{k_{\text{v}}(V - V_0)}{2V_0}$$

#### Lattice model



Shear modulus

Area compression modulus

Young's modulus

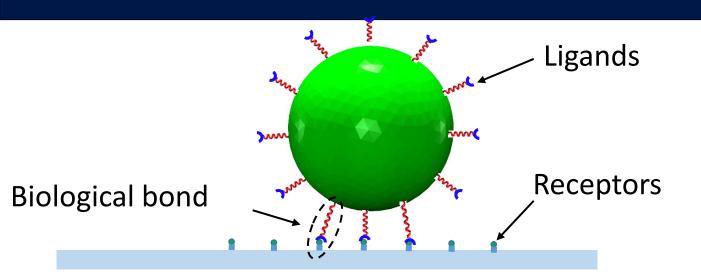
$$\mu_0 = \frac{\sqrt{3}k_B T}{4p l_m x_0} \left( \frac{x_0}{2(1-x_0)^3} - \frac{1}{4(1-x_0)^2} + \frac{1}{4} \right) + \frac{3\sqrt{3}k_p}{4l_0^3}$$

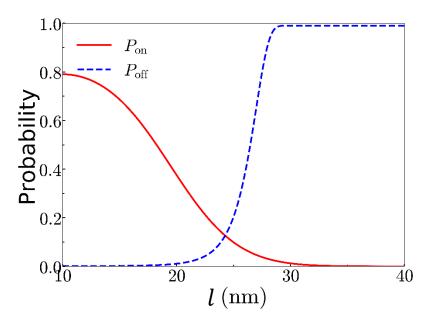
$$K = 2\mu_0 + k_a + k_d$$

$$Y = \frac{4K\mu_0}{K + \mu_0}$$

# **Ligand-Receptor Binding**

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$$F(l) = k_s(l - l_0)$$

One-to-One interaction!

$$P_{on} = \begin{cases} 1 - e^{-k_{on}\Delta t} & for \quad l < d_{on} \\ 0 & for \quad l \ge d_{on} \end{cases}$$

$$k_{on} = k_{on}^0 \exp\left(-\frac{\sigma_{on}(l-l_0)^2}{2k_B T}\right)$$

$$F(l) = k_s(l - l_0) \qquad k_{off} = k_{off}^0 \exp\left(\frac{\sigma_{off}(l - l_0)^2}{2k_B T}\right)$$

 $k_{on}$  Reaction rate

 $k_{off}$  Dissociation rate

 $\sigma_{on}$ Effective on strength

 $\sigma_{off}$ Effective off strength

Fedosov, D. A. (2010). Brown University. Hammer, D. A., & Apte, S. M. (1992). Biophysical journal

Ye, H. et al. (2017) IEEE Transactions on Nanotechnology

# Coupling: immersed boundary method (IBM) UCONN | SCHOOL OF ENGINEERING

#### 1. Advance of fluid field

$$f_i(\mathbf{x} + \mathbf{c}_i \Delta t, t + \Delta t) - f_i(\mathbf{x}, t) = -\frac{\Delta t}{\tau} \left( f_i - f_i^{\text{eq}} \right) + F_i \Delta t$$

2. Velocity interpolation

$$u_i^{\rm s}(\mathbf{X}^{\rm s},t) = \int_{\Omega} u_i(\mathbf{x},t) \delta(\mathbf{x} - \mathbf{x}^{\rm s}(\mathbf{X}^{\rm s},t)) d\Omega$$

3. Update solid field

$$U = \mu^{s}(I_1 - 3)/2 + \lambda(\ln I)^2/2 - \mu^{s} \ln I$$

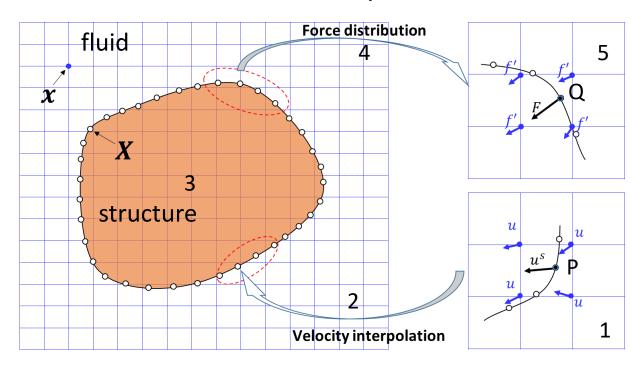
4. Force distribution

$$f_i^{FSI,s}(\mathbf{X},t) = \int_{\Omega^s} F_i^{FSI,s}(\mathbf{X}^s,t) \delta(\mathbf{X} - \mathbf{X}^s(\mathbf{X}^s,t)) d\Omega$$

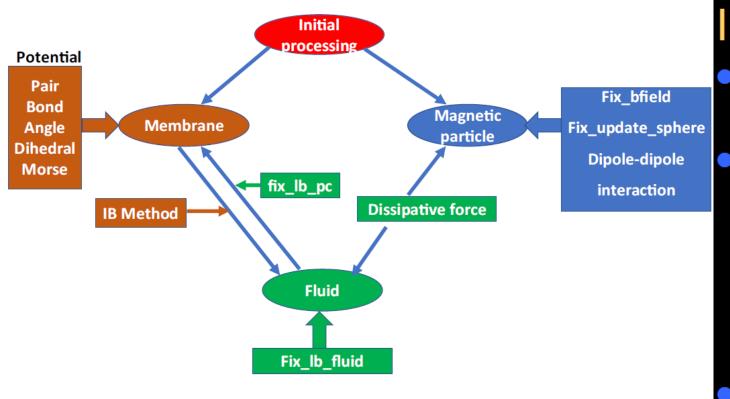
5. Advance fluid field with body force

$$f_i(\mathbf{x} + \mathbf{c}_i \Delta t, t + \Delta t) - f_i(\mathbf{x}, t) = -\frac{\Delta t}{\tau} \left( f_i - f_i^{\text{eq}} \right) + F_i \Delta t$$

#### Immersed boundary method



$$\delta(r) = \begin{cases} \frac{1}{4} \left( 1 + \cos\left(\frac{\pi|r|}{2}\right) \right), & r \le 2\\ 0, & r > 2 \end{cases}$$



# Implementation in LAMMPS:

- MPI domain decomposition taken same as particles in LAMMPS
- Implemented as fixes:
   fix lb\_fluid applies force of particle on
   fluid
   fix viscous\_lb applies force of fluid on
   particles
   along with fix momentum lb
- Hook into update.cpp : units
   Otherwise just a regular user package

**LAMMPS** is a classical molecular dynamics code with a focus on materials modeling. It's an acronym for Large-scale Atomic/Molecular Massively Parallel Simulator. LAMMPS has potentials for solid-state materials (metals, semiconductors) and soft matter (biomolecules, polymers) and coarse-grained or mesoscopic systems. It can be used to model atoms or, more generically, as a parallel particle simulator at the atomic, meso, or continuum scale.

#### Lagrangian beam

Governing equation of beam

$$U = \frac{1}{2} \sum_{i} k_{s} (dx)^{2} + \frac{1}{2} \sum_{i} k_{\theta} (d\theta)^{2}$$

#### Lagrangian plate

Governing equation of plate

$$U({x_i}) = U_{stretching} + U_{bending}$$

$$U = \mu^{s} (I_1 - 3)/2 + \lambda (\ln J)^2/2 - \mu^{s} \ln J$$

#### Lagrangian solid

Governing equation of neo-Hookean solid

$$\rho^{s} \left( \frac{\partial^{2} \mathbf{u}^{s}}{\partial t^{2}} \right) = F + f^{\text{FSI,s}}$$

$$F = \partial U/\partial X$$

$$U = \mu^{s}(I_1 - 3)/2 + \lambda(\ln J)^2/2 - \mu^{s} \ln J$$

$$u_i^{\mathrm{m}}(\mathbf{X}^m,t) = \int_{\Omega} u_i(\mathbf{x},t) \, \delta(\mathbf{x} - \mathbf{x}^m(\mathbf{X}^m,t)) \mathrm{d}\Omega$$

$$u_i^{\mathrm{S}}(\mathbf{X}^s,t) = \int_{\Omega} u_i(\mathbf{x},t) \, \delta(\mathbf{x} - \mathbf{x}^s(\mathbf{X}^s,t)) \mathrm{d}\Omega$$
Velocity interpolation

**IBM** 

LAMMPS

Incompressible fluid solver

$$(\partial_t + e_{i\alpha}\partial_\alpha)f_i = -\frac{1}{\tau}(f_i - f_i^{eq}) + f_i^{\text{FSI},m} + f_i^{\text{FSI},s}$$

**IBM** 

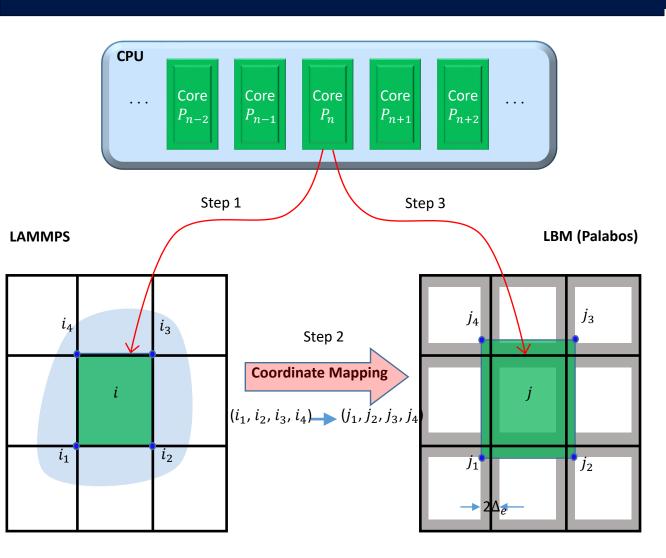
Force distribution

$$f_i^{\text{FSI},m}(\mathbf{x},t) = \int_{\Omega^S} f_i^{\text{FSI},m}(\mathbf{X}^m,t) \,\delta(\mathbf{x} - \mathbf{x}^m(\mathbf{X}^m,t)) d\Omega$$

$$f_i^{\text{FSI},s}(\mathbf{x},t) = \int_{\Omega^s} f_i^{\text{FSI},s}(\mathbf{X}^s,t) \,\delta(\mathbf{x} - \mathbf{x}^s(\mathbf{X}^s,t)) \,\mathrm{d}\Omega$$

### Implementation: Parallel computation

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CPU mapping to complete velocity interpolation and force spreading

#### Algorithm 1: Immersed boundary-based spatial decomposition

#### **Run LAMMPS**

Read simulation box  $\Gamma$ 

Partition  $\Gamma$  into N regular bricks #comm brick

**Run interface** #contained in file: *latticeDecomposition.hh* 

Pass bricks information to Palabos

#### Run Palabos

Create N blocks with the information from LAMMPS

For loop m in N blocks

Assign block information to processor numbered n

End loop

Initialization parallelism in Palabos

#### Algorithm 2: Data communication between LAMMPS and Palabos

#### Run LAMMPS

Obtain coordinates of structure nodes x(X, t), and pass them to IB interface.

#### **Run Palabos**

Obtain the velocity of fluid nodes u(x, t), and pass them to IB interface.

**Run IB interface** #contained in files: *ibm2D.hh* or *ibm3D.hh* 

Interpolate the velocity  $u^s$  at location x(X,t), and return them to LAMMPS.

#### **Run LAMMPS**

Calculate the FSI force F, and pass them to IB interface

**Run IB interface** #contained in files: *ibm2D.hh* or *ibm3D.hh* 

Spread FSI force F to the fluid nodes f' at location (x, t), and pass them to Palabos.

#### **Run Palabos**

Use the body force nodes f' to advance fluid field.

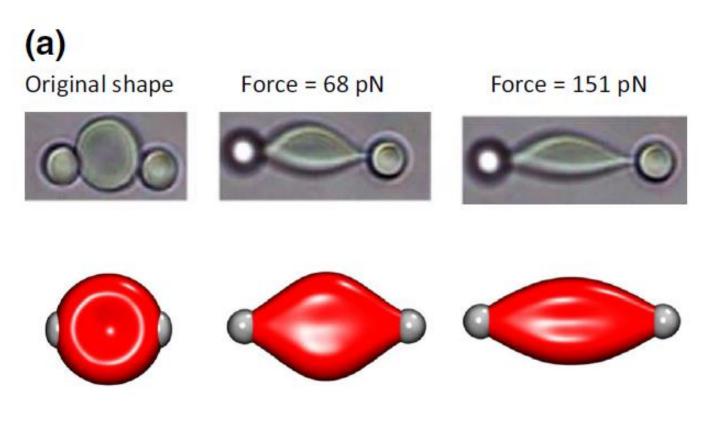
#### **Run LAMMPS**

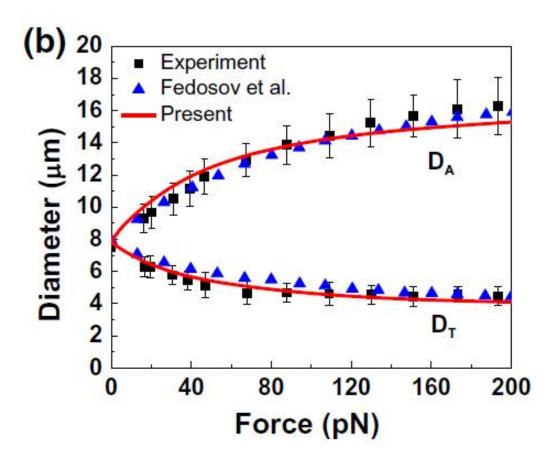
Use FSI force **F** to update the new location of structure.

Ye, H. et al. (2020) Computer Physics Communications



#### Stretching of a single RBC

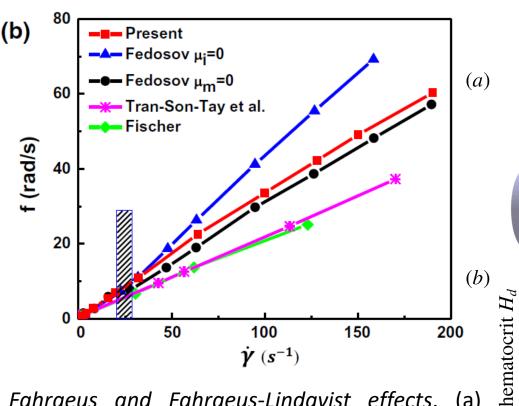




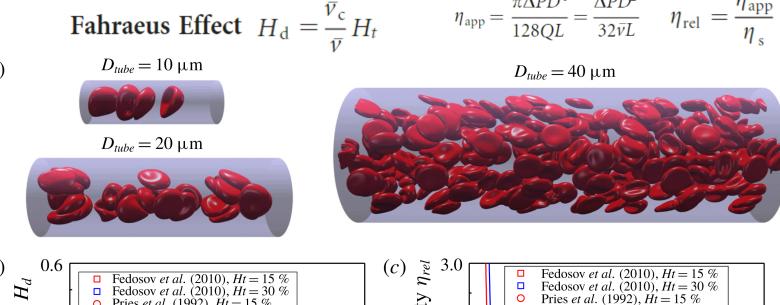
Stretching test of a single RBC. **a** Configuration of stretched RBC with different applied forces. **b** Diameters of RBC along stretching direction and transverse direction, denoted by  $D_A$  and  $D_T$ , respectively. The experimental results are reproduced with permission from Suresh S et al. Acta Biomater 23:S3–S15, 2015.

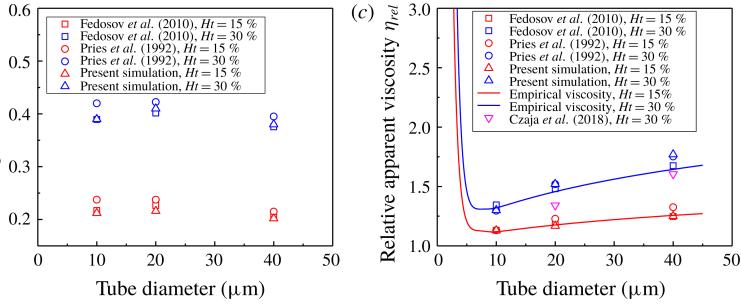
#### Tank-treading and tumbling a single RBC in simple shear flow

#### Fahraeus-Lindqvist Effect



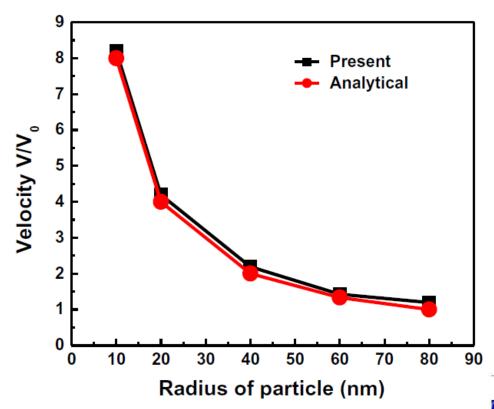
Fahraeus and Fahraeus-Lindqvist effects. (a) Snapshots to show the tube flow with different diameters under hematocrit 15%. (b) Fahraeus effect: discharge hematocrit comparison. (c) Fahraeus-Lindqvist effect: relative viscosity validation.





# Validation-Magnetic Particle in Flow

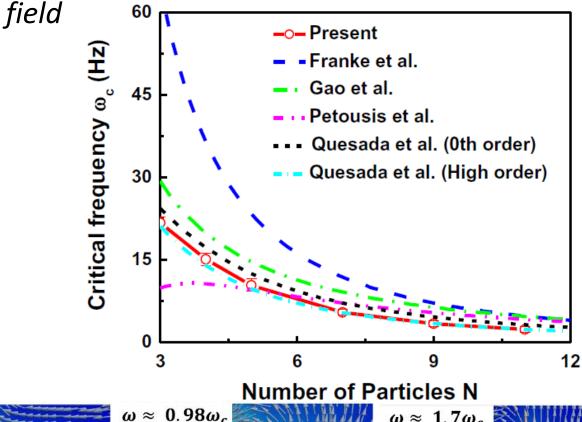
Moving of a single magnetic particle under non-uniform external magnetic field

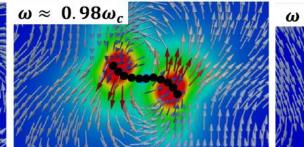


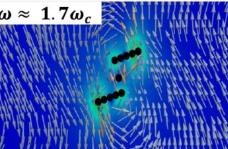
$$V_i = \frac{F_i^B}{6\pi\,\mu a}$$

 $\omega \approx 0.2\omega_c$ 

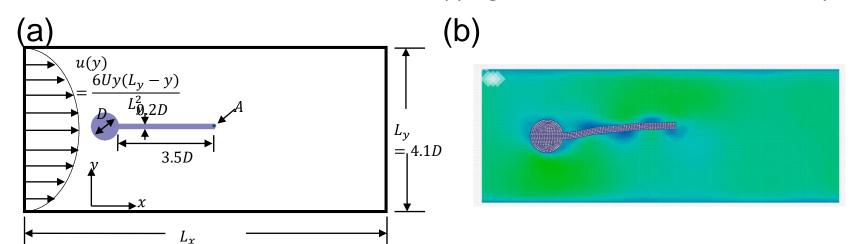
Breakup of magnetic chain in rotating magnetic







Flow-induced flapping of an elastic 2D beam behind a cylinder

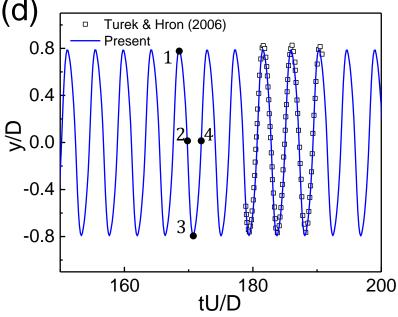


$$St = fU/D$$
 Oscillating behavior

$$C_D = F_{\chi}/(0.5\rho U^2 D)$$
 Drag coefficient

(c)		(d) Figure Turek &		
(0)	□ Turek & Hron (2006)			
0.0	Present	$0.8 \begin{array}{ c c c c c c c c c c c c c c c c c c c$		
-0.1 \(\frac{1}{\times}\) -0.2 -0.3		0.4 0.4 0.0 -0.4 -0.8		
	160 180 20	0 160		
tU/D				

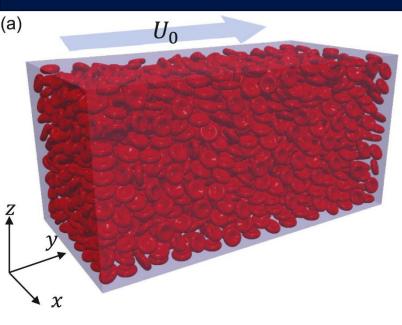
= 11D

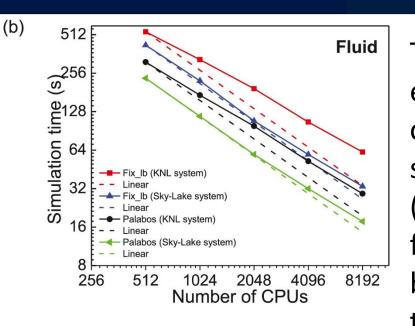


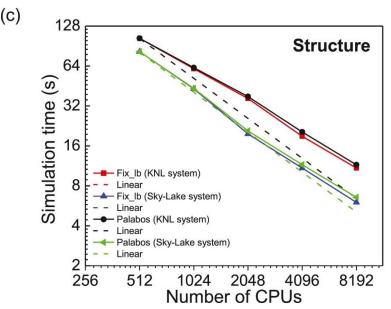
Works	Amplitude	Strouhal number	Drag coefficient
Turek and Hron (2006)	0.83	0.19	4.13
Tian et al. (2014)	0.78	0.19	4.11
Lin et al. (2019)	0.81	0.19	4.10
Present	0.79	0.19	4.10

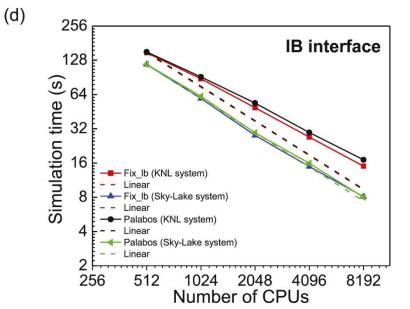
# **Scalability of Computational Framework**

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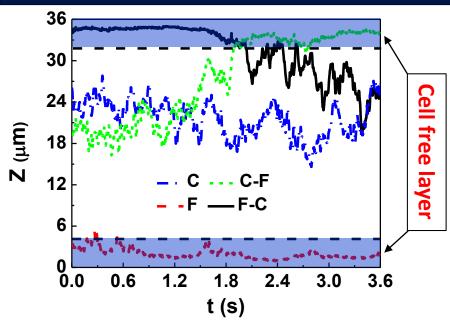


The computational efficiency is examined by estimating the time consumption used to run 2000 steps within present program. (a) Model of large-scale blood flow simulations with 2000 red blood cells. Separate simulation times of (b) fluid part, structure part, and interface part for running time steps in different systems with different solvers.

Huilin Ye et al. Computer Physics Communications Volume 256, November 2020, 107463

# **Margination of Drug Carriers**

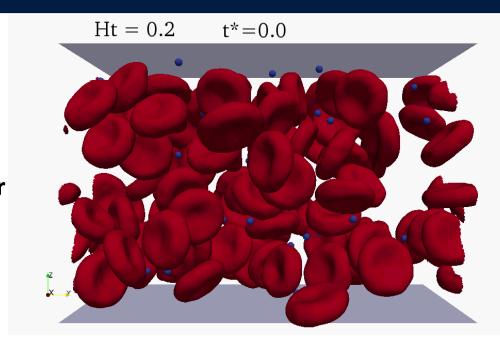
# UCONN | SCHOOL OF ENGINEERING



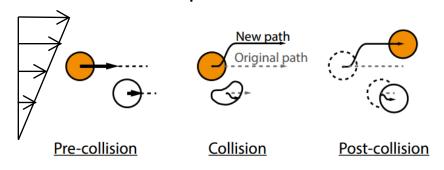
### Four types of motion

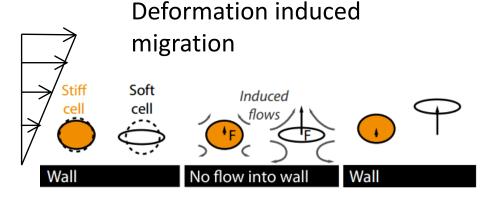
- (A) C: stay in center region
- (B) F: stay in cell free layer
- (C) C-F: marginate from center to cell free layer
- (D) F-C: demarginate from cell free layer to center region

$$\Pi = \frac{n_{\rm f}(t) - n_{\rm f}(0)}{N - n_{\rm f}(0)}$$



Particle-particle collisions





☐ Anomalous vascular dynamics of nanoworms within blood Flow

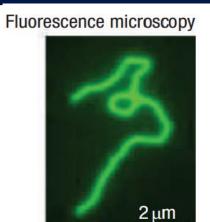
Passive targeting: Size, Shape and Stiffness

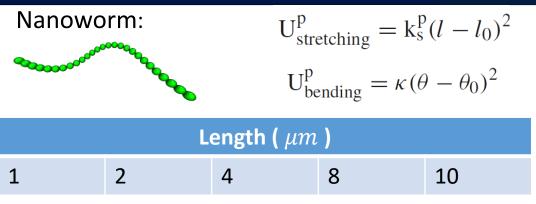
☐ Adhesion effect on margination of elastic micro-particles

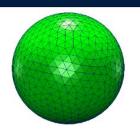
☐ Motion of particle under external magnetic field

# Circulation: Vascular dynamics of nanoworms in blood flow

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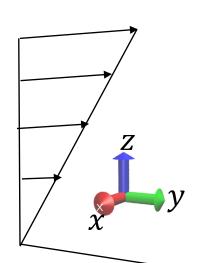




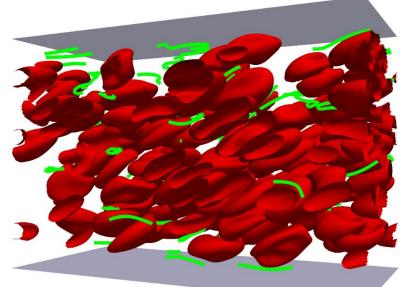


Sphere: 1  $\mu m \ (N_p = 50)$ 

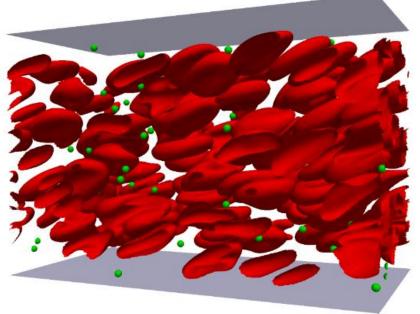
$$\mathbf{u} = \{0, \ \gamma \dot{z}, \ 0\} \quad \dot{\gamma} = 248 \ \mathrm{s}^{-1}$$



$$L_x \times L_y \times L_z = 27 \mu m \times 54 \mu m \times 36 \mu m$$



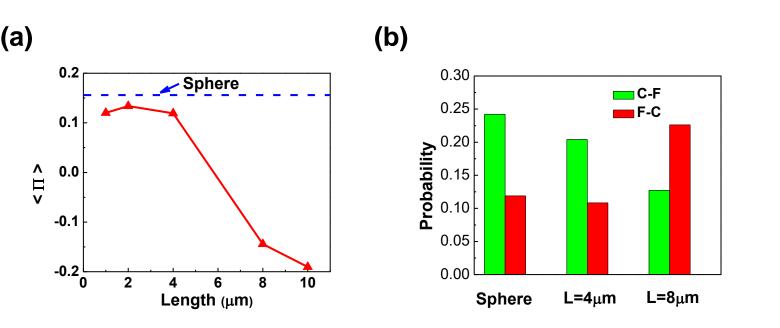
Ht: 20% ( $N_R$ = 108)



Geng, Y. et al. (2007) Nature nanotechnology

# **Demargination of Semiflexible Nanoworms**



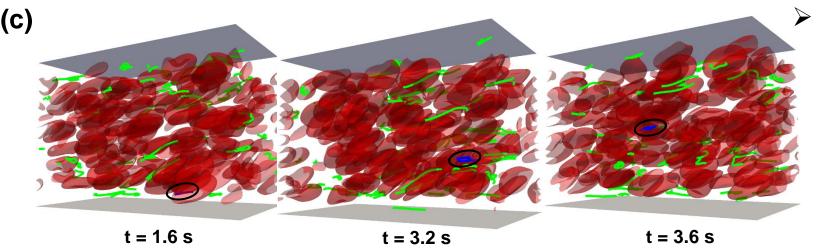


#### ☐ Results

- Short nanoworms show margination behavior
- Margination probability of long nanoworms are negative

#### □ Reasons

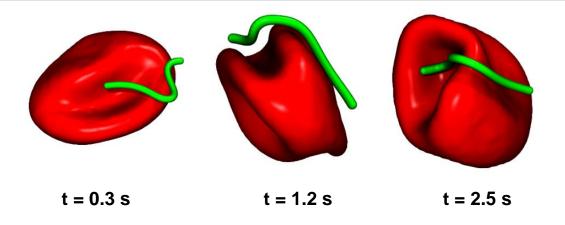
- C-F motion dominates for short nanoworms
- F-C motion dominates for long nanoworms. Demargination outperforms margination.

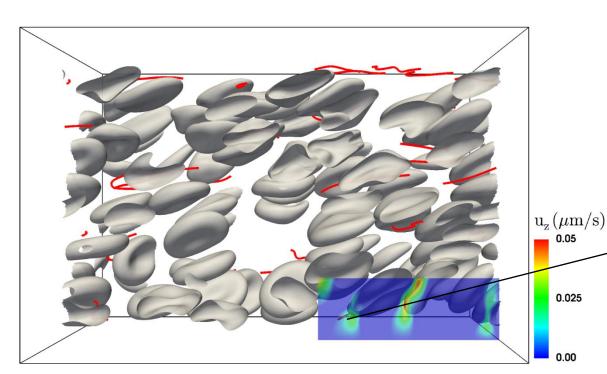


Ye, Huilin, et al. ACS Biomaterials Science & Engineering (2017).

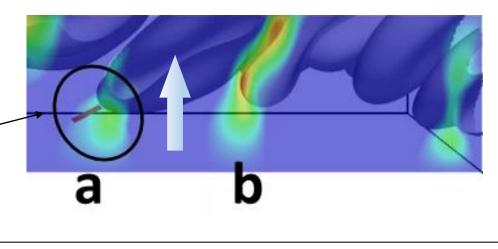
# **Physical Mechanisms**

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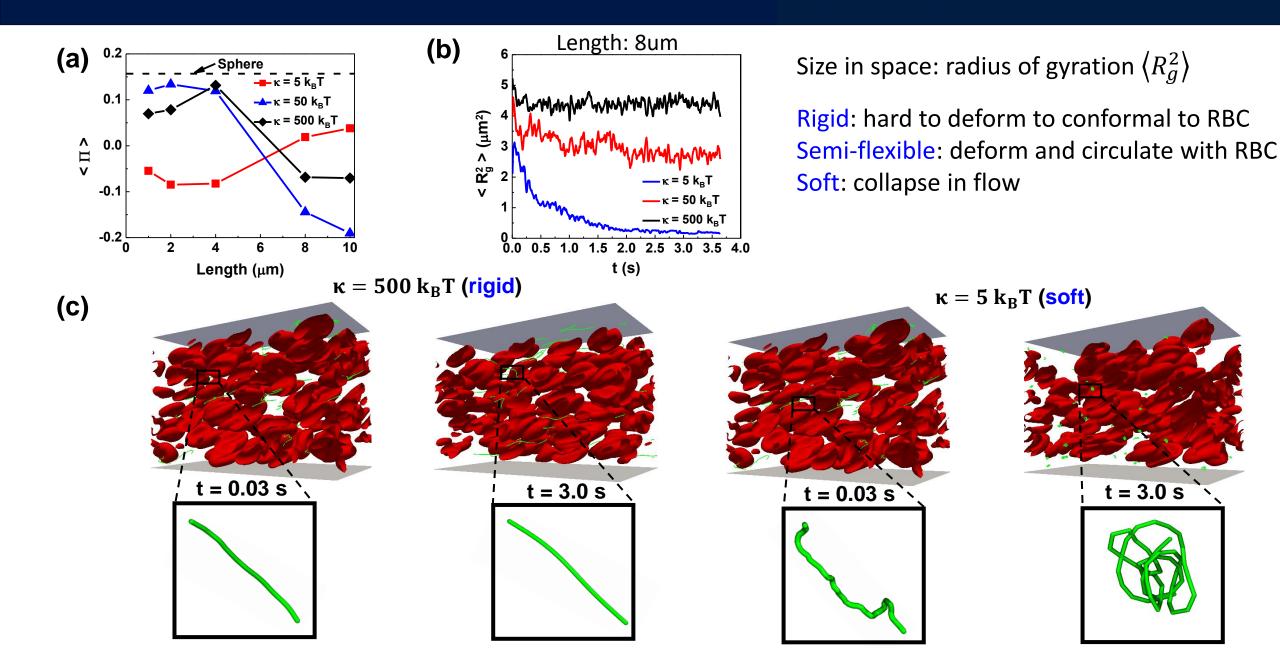
- ☐ Low Margination: Flexible nanoworms deform to the same shape with RBCs and intertwine with them, moving along bulk flow.
- ☐ Demargination: Perturbation of fluid flow around RBCs due to deformation induces lateral flow, and drag the nanoworms to the center of channel.



Ye, Huilin, et al. ACS Biomaterials Science & Engineering (2017).

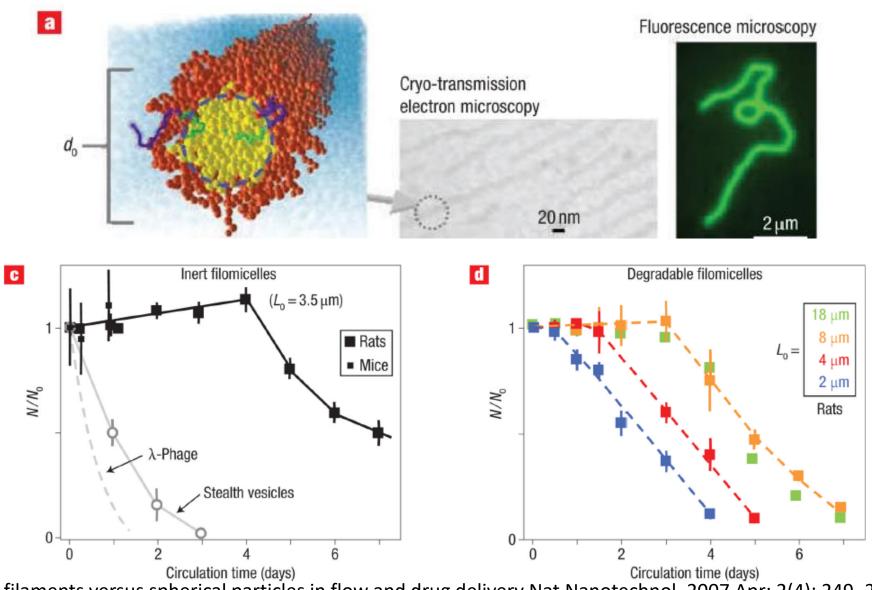
### **Stiffness effect**

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## **Experimental Observation**





Shape effects of filaments versus spherical particles in flow and drug delivery Nat Nanotechnol. 2007 Apr; 2(4): 249–255.

☐ Anomalous vascular dynamics of nanoworms within blood Flow

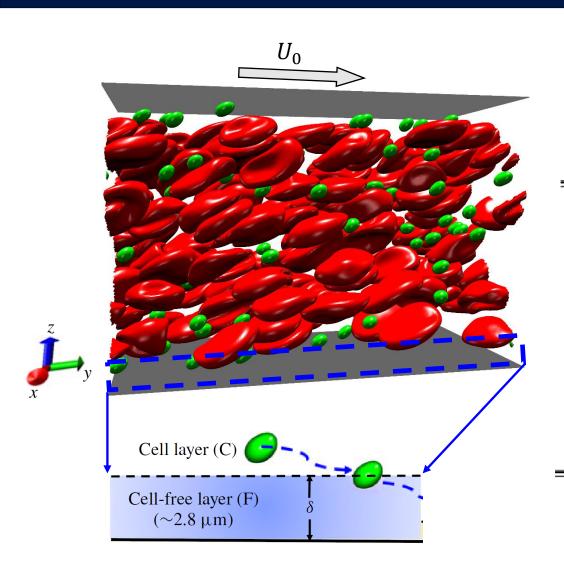
☐ Adhesion effect on margination of elastic micro-particles

Passive targeting: Stiffness and Adhesion (Surface)

☐ Motion of particle under external magnetic field

## **Margination and Adhesion**

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$$L_z \times L_z \times L_z = 27 \times 54 \times 36 \ \mu m^3, \ \gamma = \frac{U_0}{L_z} = 200 \ s^{-1}$$

162 RBCs and 80 particles Volume fraction of RBC: 30%

#### **Dimensionless numbers**

capillary number  $Ca = \mu \dot{\gamma} R/\mu_0$ 

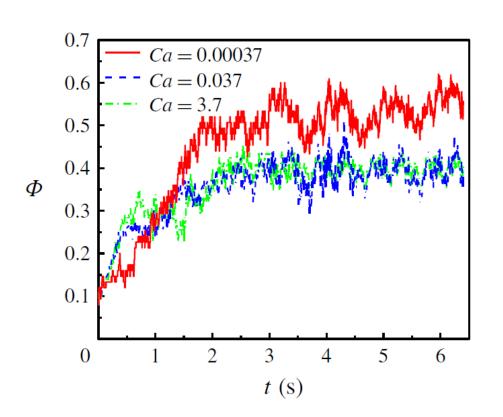
Rigid Soft

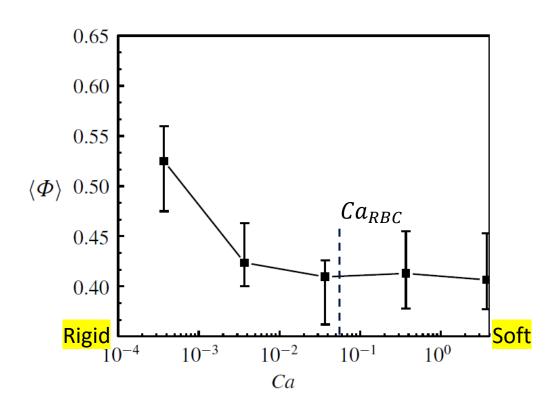
adhesion number  $Ad = k_s/\mu \dot{\gamma} R$ 

Weak Strong

#### **Margination Probability**

$$\Phi(t) = \frac{n_f(t)}{N}$$

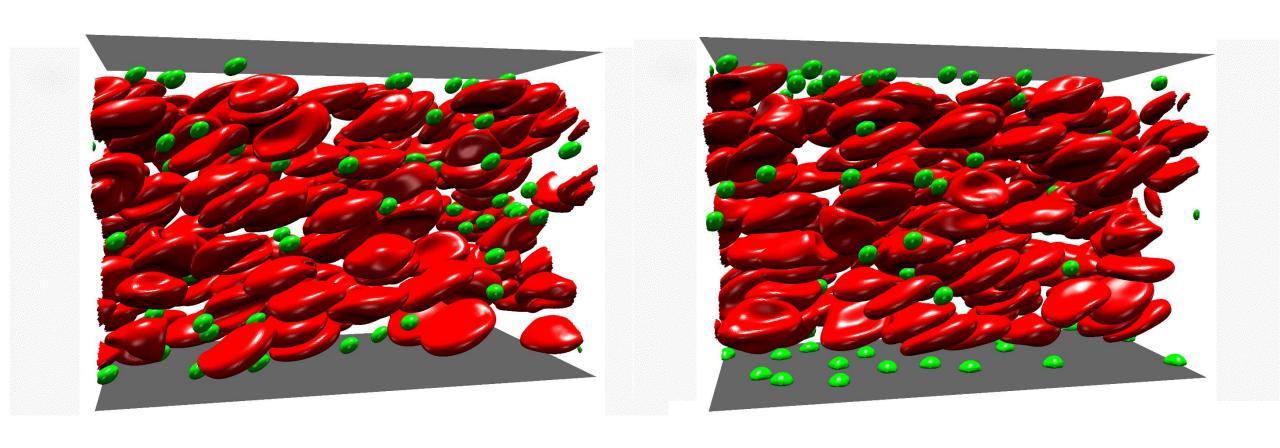




- Rigid particle has stronger margination (consistent with Kumar & Graham, 2011)
- Margination is independent on the stiffness when particle becomes soft enough

# Interplay effect

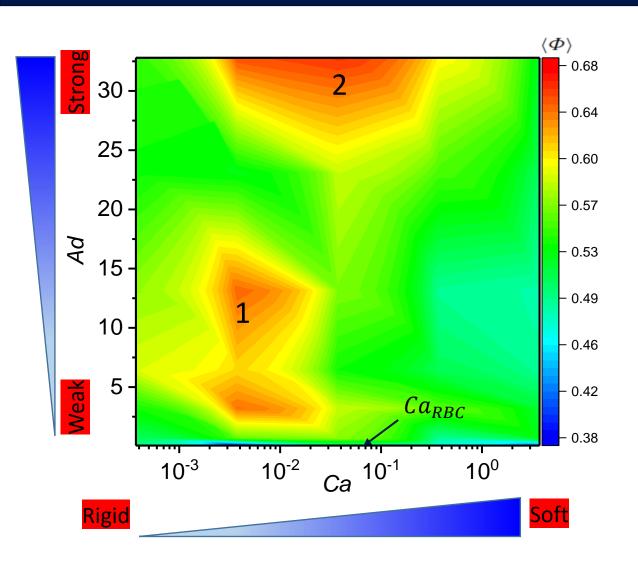




No adhesion With adhesion

# Interplay effect: two optimal regions





#### Two optimal regions (high probability)

- (1) large stiffness and weak adhesion;
- (2) Moderate stiffness and strong adhesion

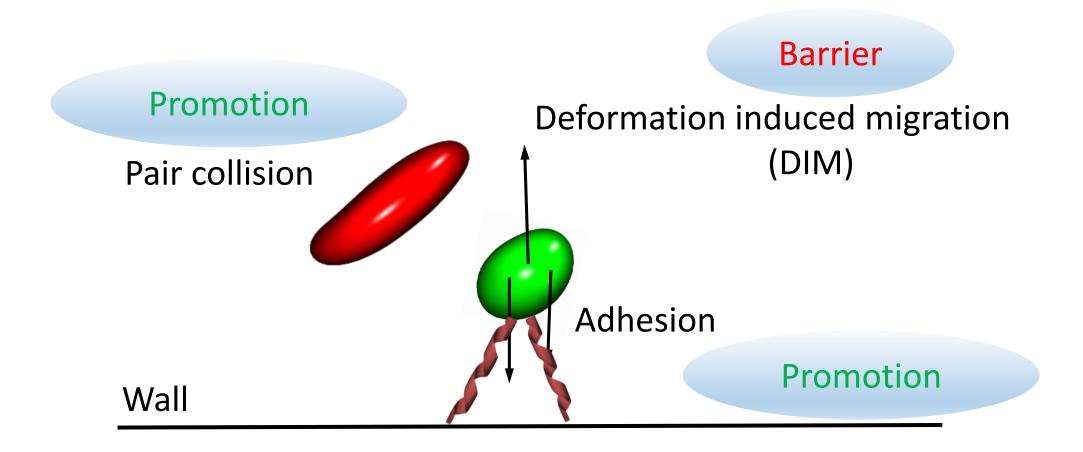
#### Interplay of stiffness and adhesion

- (1) Rigid or softer particle is not always better
- (2) Stronger adhesion is also not better

What's the underlying mechanism?

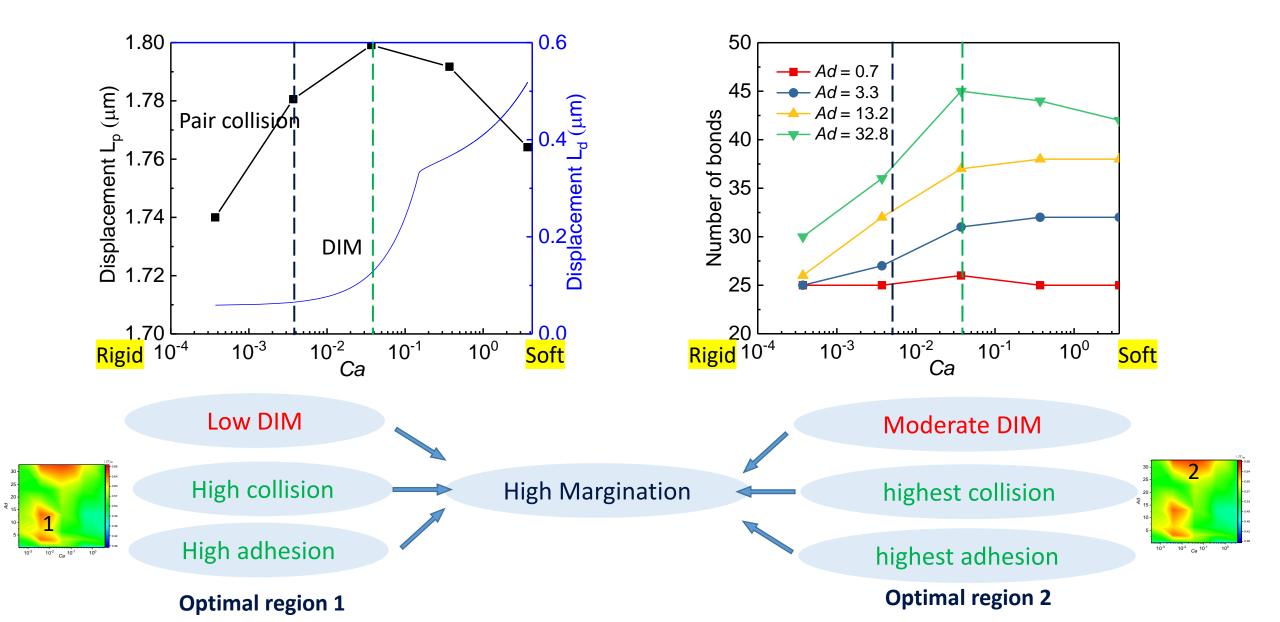
# Interplay effect: underlying mechanism





## Interplay effect: interpretation





☐ Anomalous vascular dynamics of nanoworms within blood Flow

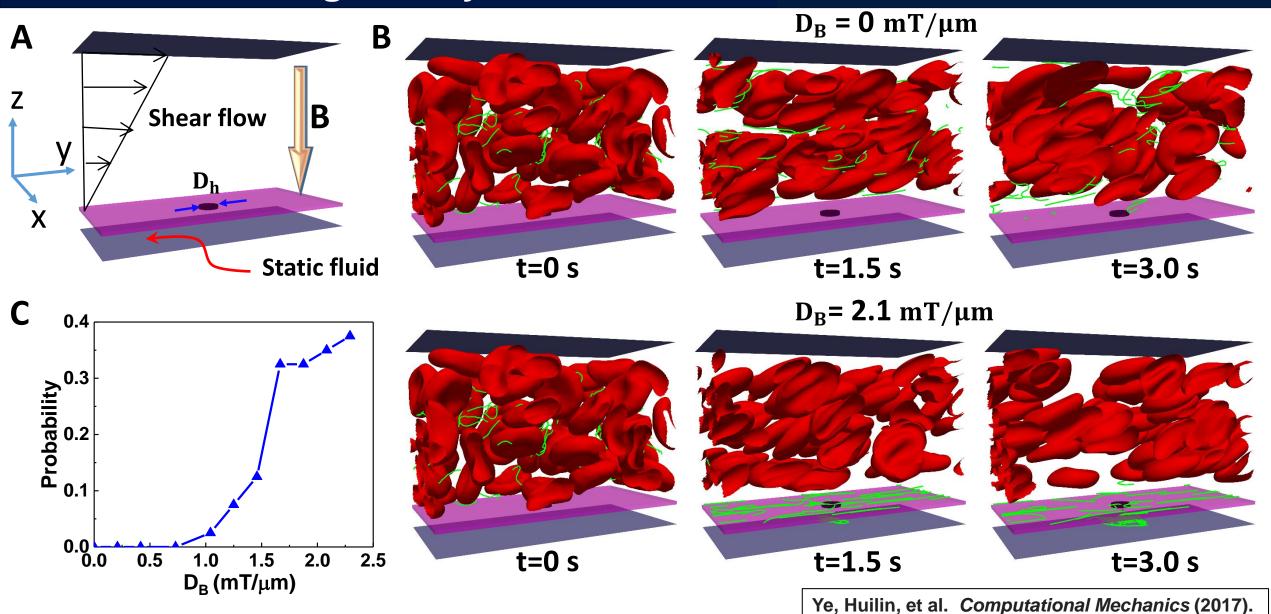
☐ Adhesion effect on margination of elastic micro-particles

☐ Motion of particle under external magnetic field

**Active targeting: External triggering** 

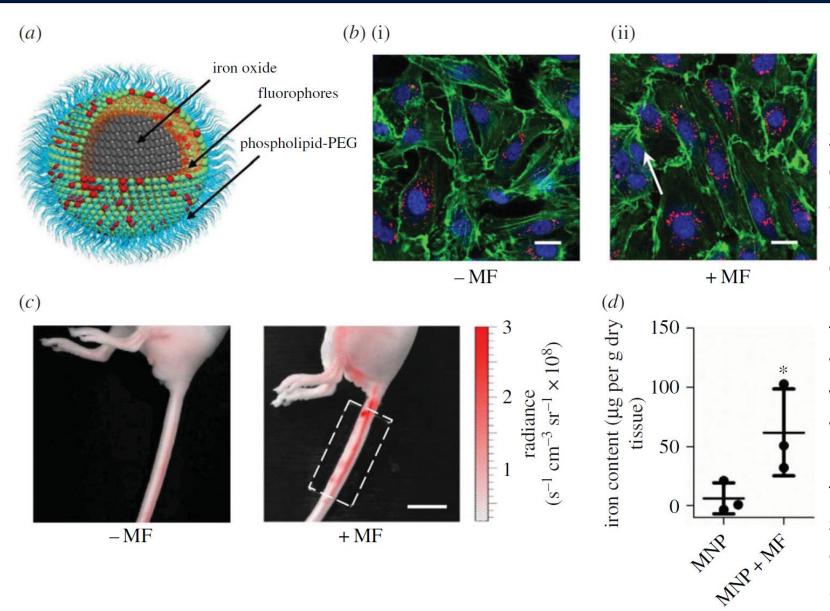
# Magnetic Guidance of Particle Penetration through Leaky Wall





### **Experimental Observation**

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Experimental results for enhanced permeability of vascular endothelium an externalmagnetic field. (a) Schematic diagram of magnetic particle containing a magnetite nanocrystal and phospholipid-polyethylene glycol (PEG) coating. (b) Endothelial cells containing particles magnetic incubated without and with magnetic field. (c) Particles with magnetism were injected into a mouse lateral tail vein, and the opposite lateral tail vein placed under the external was magnetic field. (d) Magnetic particles in the mouse tails quantified by a superconducting quantum interference device. Qiu et al. 2017 Nat. Commun. 8, 15594.

## **Conclusion and Ongoing work**



#### **Conclusion:**

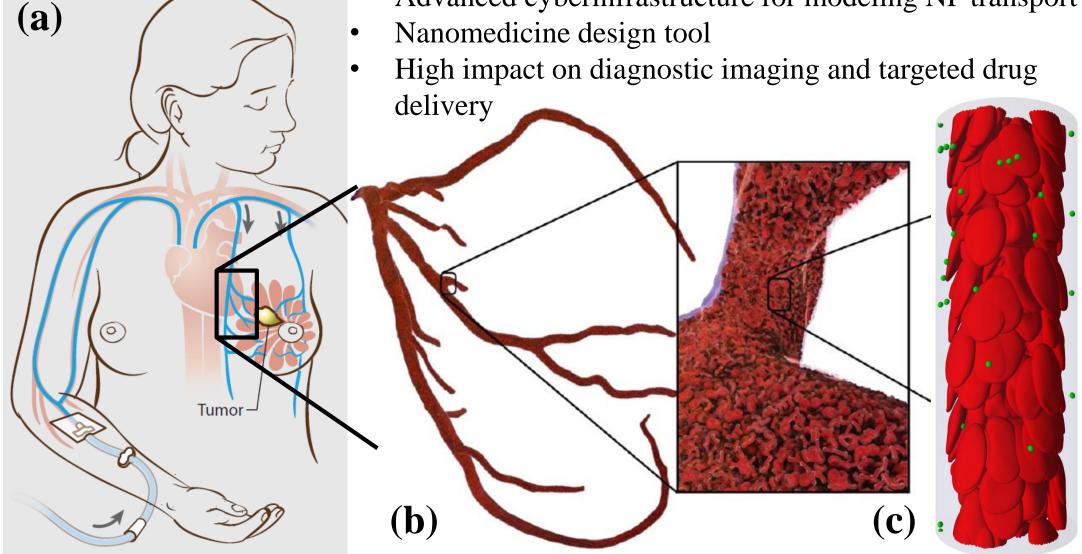
- □Filamentous nanoworms can circulate longer than spherical particle in blood flow
- ☐ Margination of elastic particles: softer is not always better
- □ External triggering magnetic field can guide the accumulation of particle at near wall and promote the permeation through leaky wall

## **Ongoing work:**

- □ Lattice Boltzmann method → complex vascular network modeling with machine learning technique (Graph Neural Network, GNN)
- □GPU-accelerated blood flow simulations (multi-platform HPCs)

#### **Anticipated outcome**

Advanced cyberinfrastructure for modeling NP transport



#### **Future Work-KOKKOS**



#### Goals: Portable, Performant, and Usable (https://github.com/kokkos)

- ☐ Portable to Advanced Manycore Architectures
  - Multicore CPU, NVidia GPU, Intel Xeon Phi (potential: AMD Fusion)
  - Maximize amount of user (application/library) code that can be compiled without modification and run on these architectures
  - Minimize amount of architecture-specific knowledge that a user is required to have
  - Allow architecture-specific tuning to easily co-exist
  - Only require C++1998 standard compliant

#### ☐ Performant

- Portable user code performs as well as architecture-specific code
- Thread scalable not just thread safety (no locking!)

#### ☐ Usable

- Small, straight-forward application programmer interface (API)
- Constraint: don't compromise portability and performance

Edwards, H. Carter, Christian R. Trott, and Daniel Sunderland. "Kokkos: Enabling manycore performance portability through polymorphic memory access patterns." *Journal of Parallel and Distributed Computing* 74, no. 12 (2014): 3202-3216.

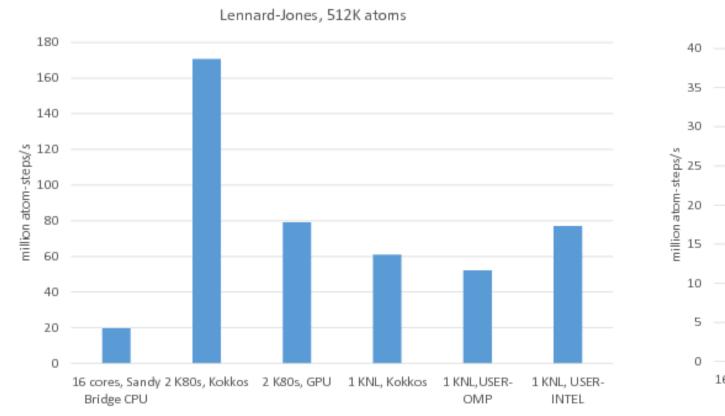
#### **Future Work-KOKKOS in LAMMPS**

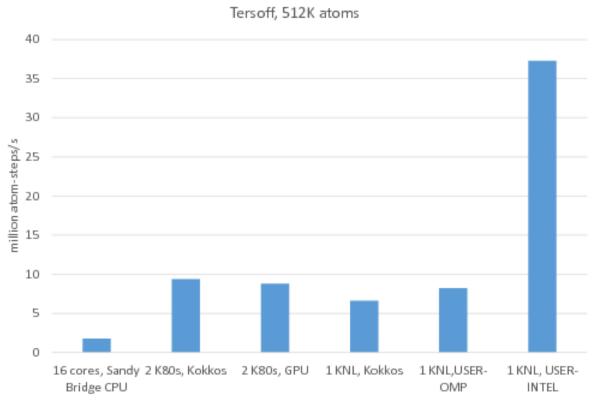


#### Accelerator benchmarks for CPU, GPU, KNL for LAMMPS

Benchmark of the Lennard-Jones potential

Benchmark of the Tersoff potential



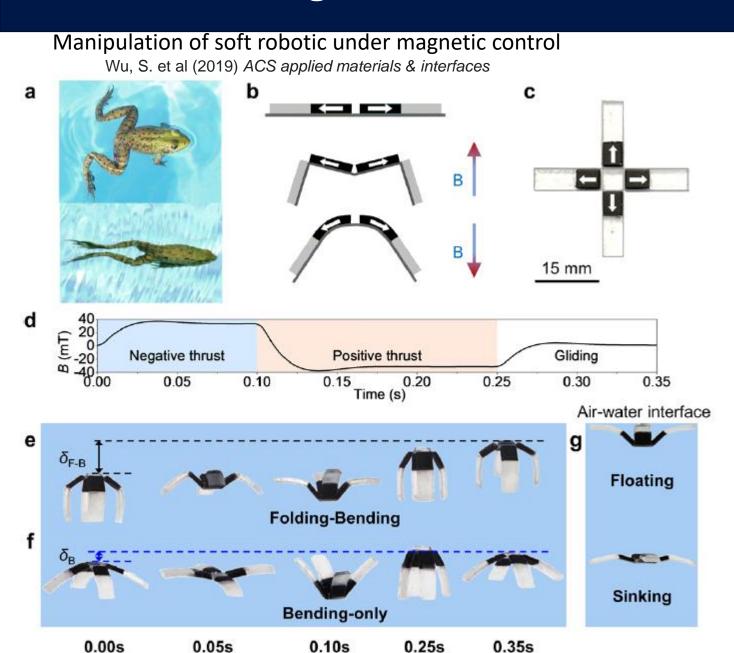


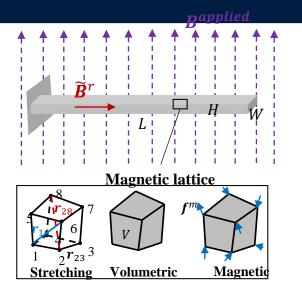
16 cores, Sandy Bridge CPU: no accelerator package, 1MPI task per core = 16 MPI tasks total

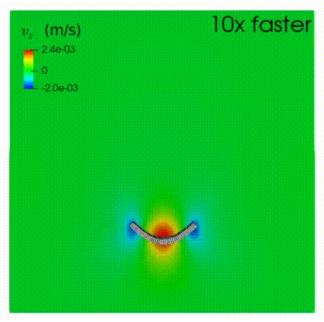
- 2 K80 GPUs, Kokkos: Kokkos package used 1 MPI task per internal K80 GPU = 4 MPI tasks total, running on 4 Sandy Bridge CPU cores, half neighbor list, newton on
- 2 K80 GPUs, GPU: GPU package used 4 MPI tasks per internal K80 GPU = 16 MPI tasks total, running on 16 Sandy Bridge CPU cores
- 1 KNL, Kokkos: Kokkos package used 64 MPI tasks x 4 hyperthreads x 4 OpenMP threads, half neighbor list, newton on
- 1 KNL, USER-OMP: USER-OMP package used 64 MPI tasks x 4 hyperthreads x 4 OpenMP threads
- 1 KNL, USER-INTEL: USER-INTEL package used 64 MPI tasks x 4 hyperthreads x 4 OpenMP threads

# **Future Work: magnetic soft robotic**

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Ye, H., Li, Y., & Zhang, T. (2021). Soft Matter, 17(13), 3560-3568.

#### **Publications**



- ☐ Ye, H., Shen, Z., & Li, Y. (2021). Adhesive rolling of nanoparticles in a lateral flow inspired from diagnostics of COVID-19. *Extreme Mechanics Letters*, 44, 101239. ☐ Ye, H., Shen, Z., Xian, W., Zhang, T., Tang, S., & Li, Y. (2020). OpenFSI: A highly efficient and portable fluid structure simulation package based on immersed-boundary method. Computer Physics Communications, 256. 107463. ☐ Ye, H., Shen, Z., Wei, M., & Li, Y. (2021). Red blood cell hitchhiking enhances the accumulation of nano-and micro-particles in the constriction of a stenosed microvessel. **Soft Matter**, 17(1), 40-56. ☐ Ye, H., Shen, Z., & Li, Y. (2019). Interplay of deformability and adhesion on margination of elastic microparticles in blood flow. Journal of Fluid Mechanics ☐ Ye, H., Shen, Z., & Li, Y. (2018). Shape dependent transport of micro-particles in blood flow: from margination to adhesion. Journal of Engineering Mechanics (Best Paper Selected by EMI FDTC) ☐ Ye, H., Shen, Z., Yu, L., Wei, M., & Li, Y. (2018). Manipulating nanoparticle transport within blood flow through external forces: an exemplar of mechanics in nanomedicine. *Proc. R. Soc. A*, 474(2211), 20170845 ☐ Ye, H., Shen, Z., & Li, Y. (2017). Cell Stiffness Governs its Adhesion Dynamics on Substrate under Shear Flow. *IEEE Transactions on Nanotechnology*
- ☐ Ye, H., Shen, Z., Yu, L., Wei, M., & Li, Y. (2017). Anomalous Vascular Dynamics of Nanoworms within Blood Flow. *ACS Biomater. Sci. Eng.*

through LAMMPS. Computational Mechanics (2017): 1-20.

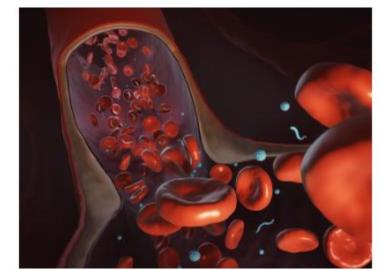
☐ Ye, H., Shen, Z., & Li, Y. (2017). Computational Modeling of Magnetic Particle Margination within Blood Flow

# Thank You!

#### TARGETING TUMORS WITH NANOWORMS

Published on April 28, 2021 by Aaron Dubrow

University of Connecticut engineer uses supercomputers and AI to improve delivery of nanomedicines for cancer







https://www.tacc.utexas.edu/-/targeting-tumors-with-nanoworms

